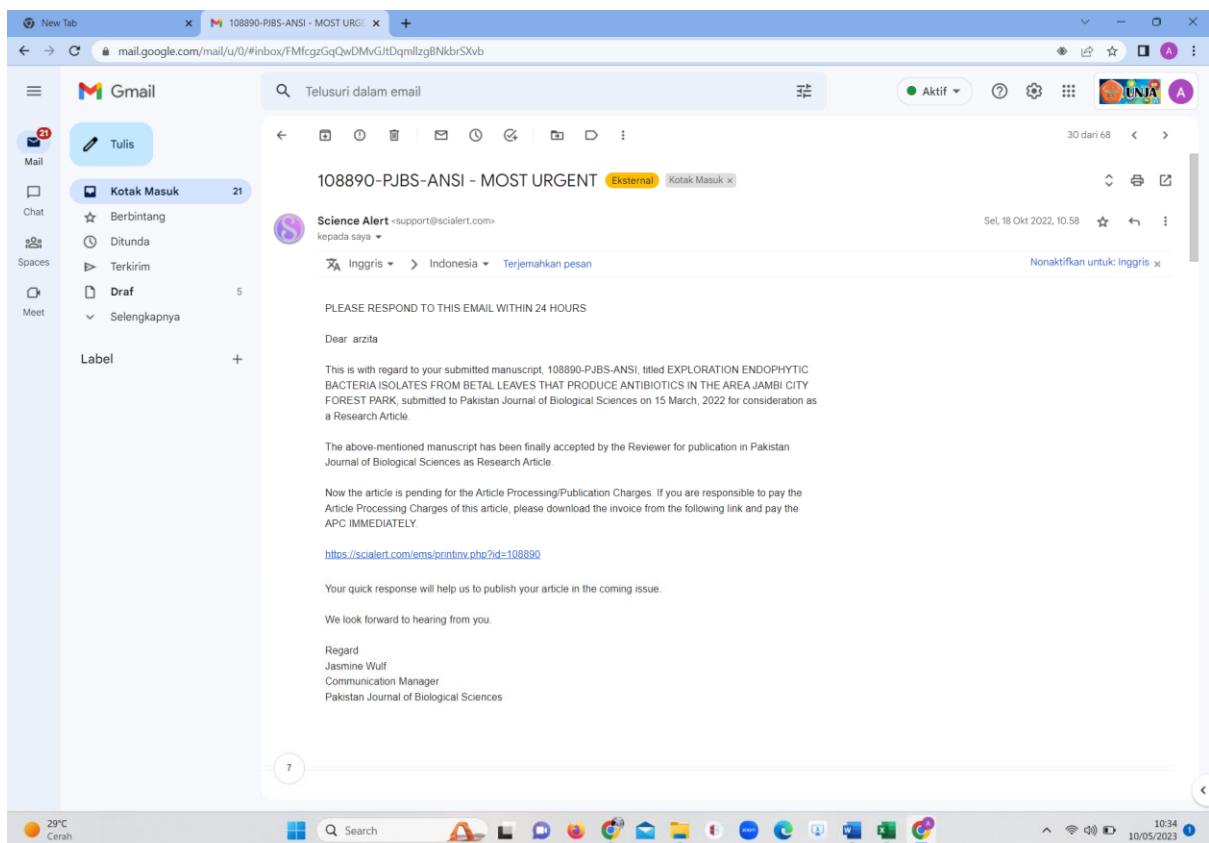
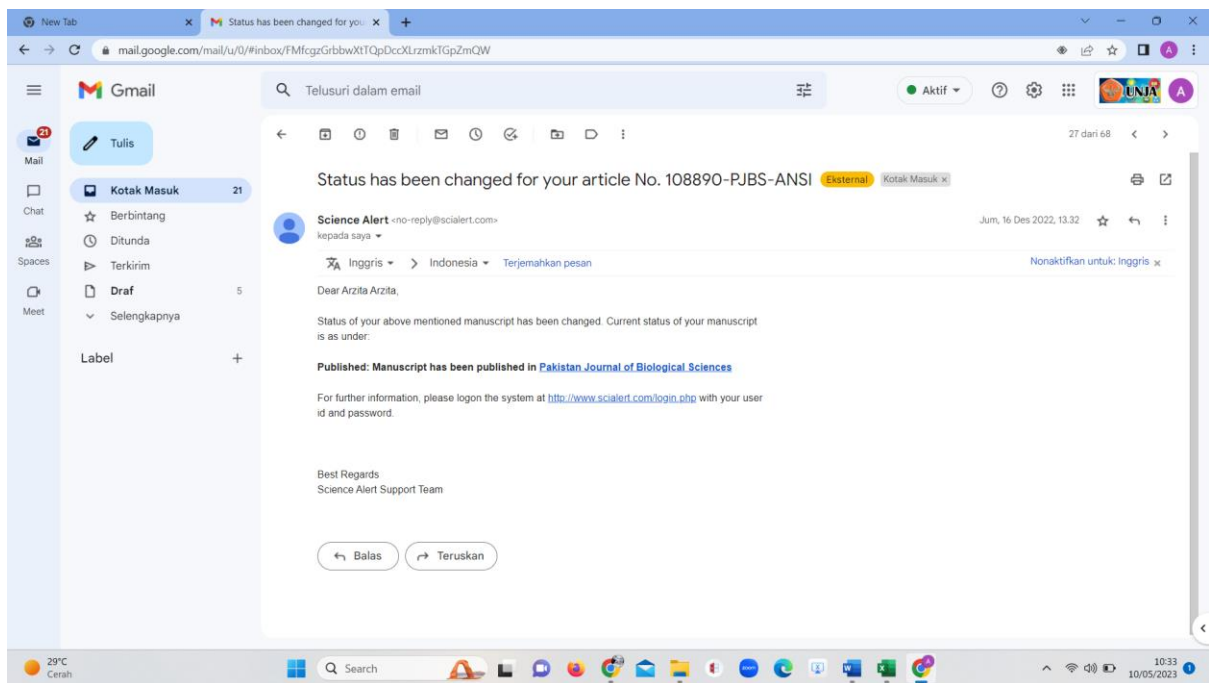
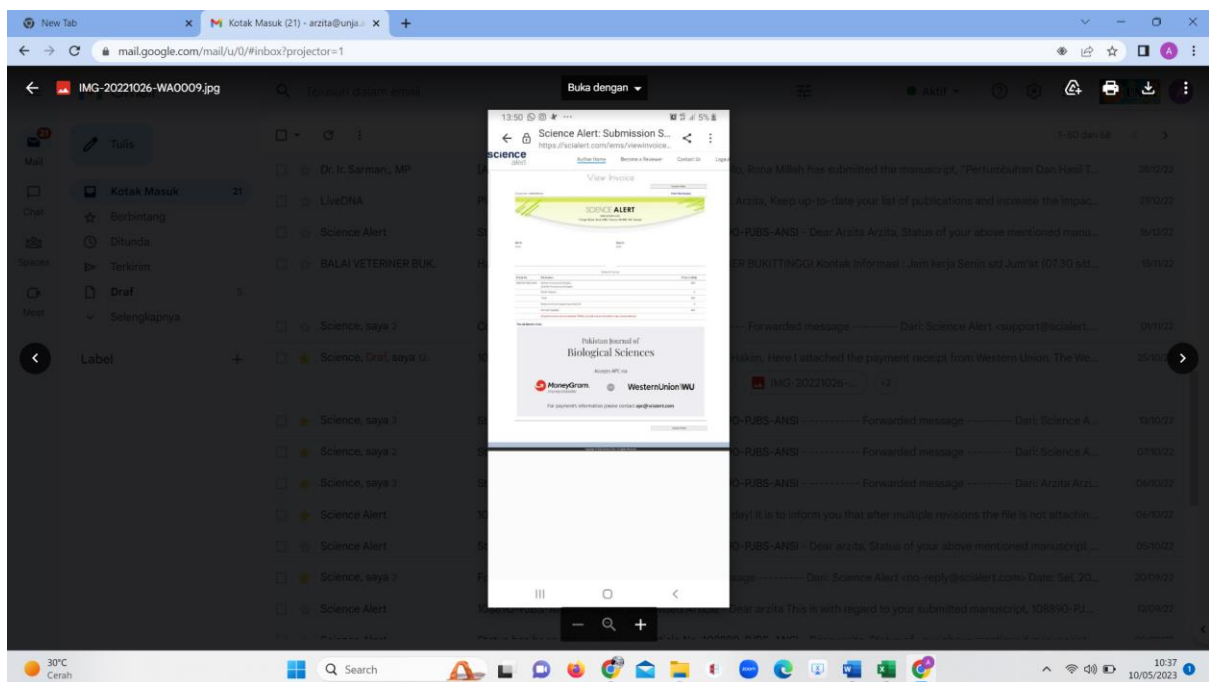
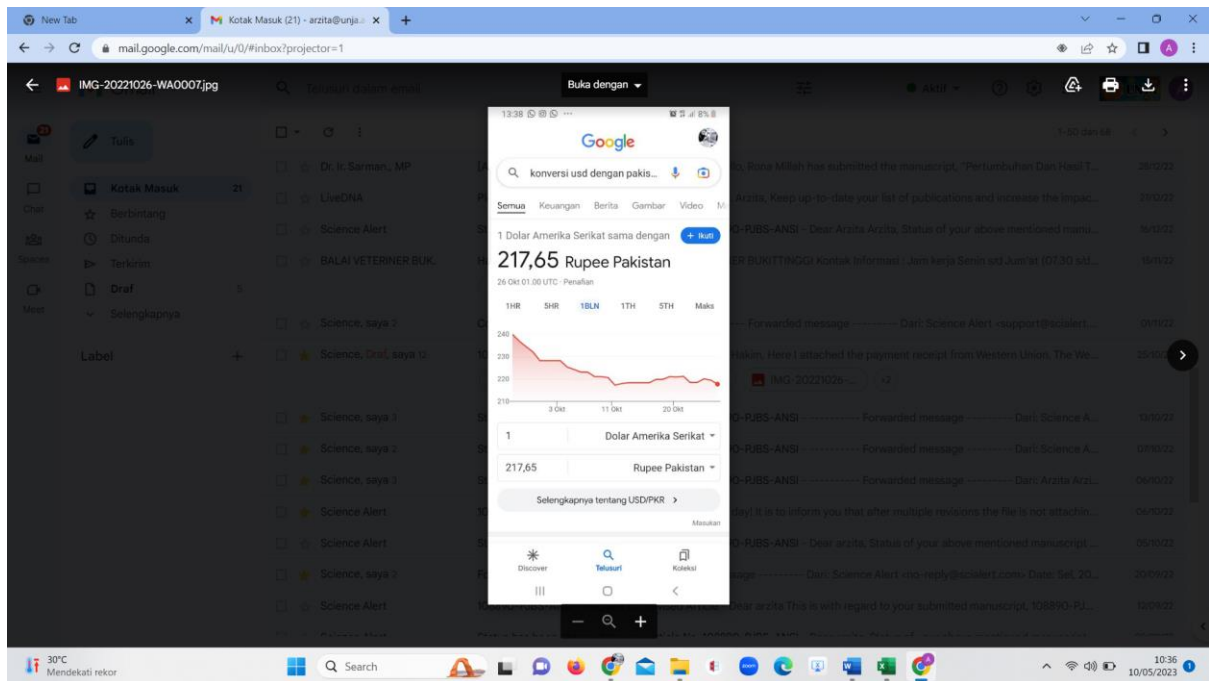
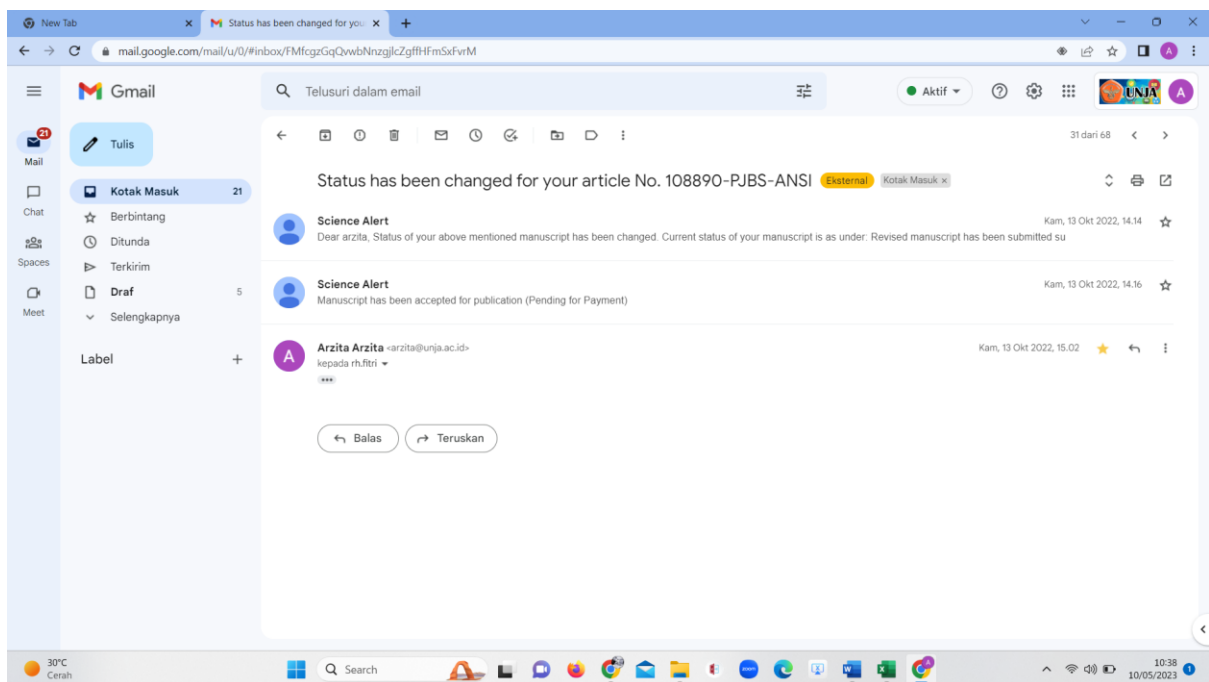
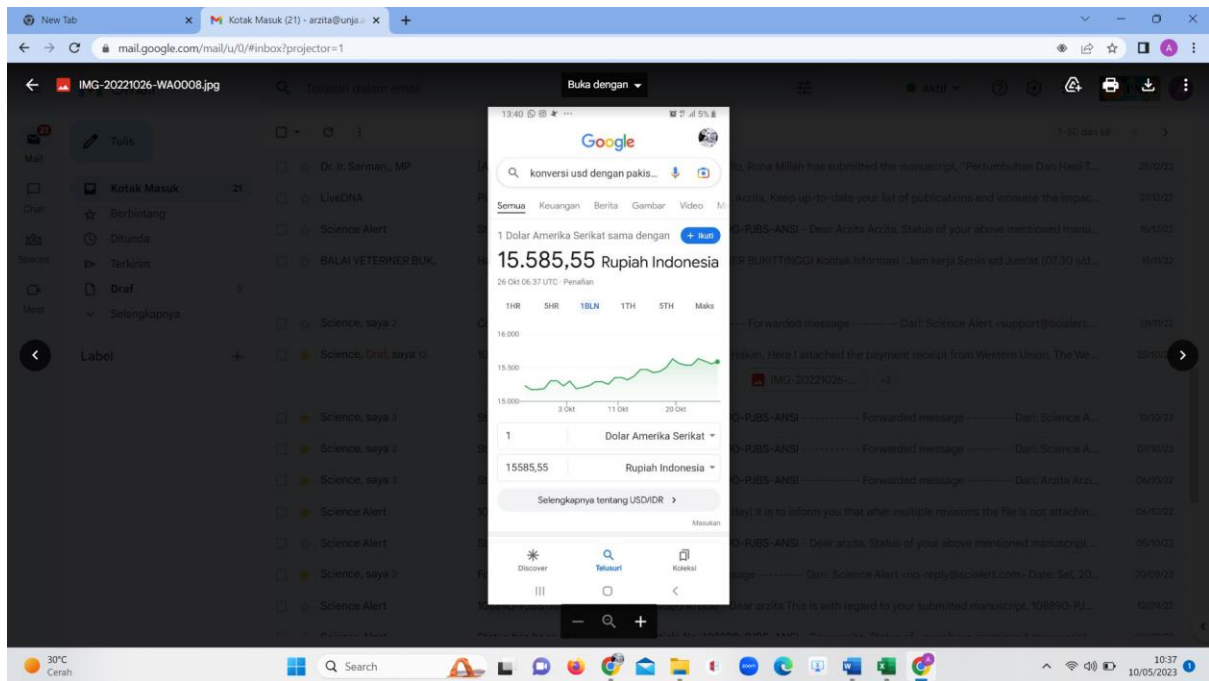
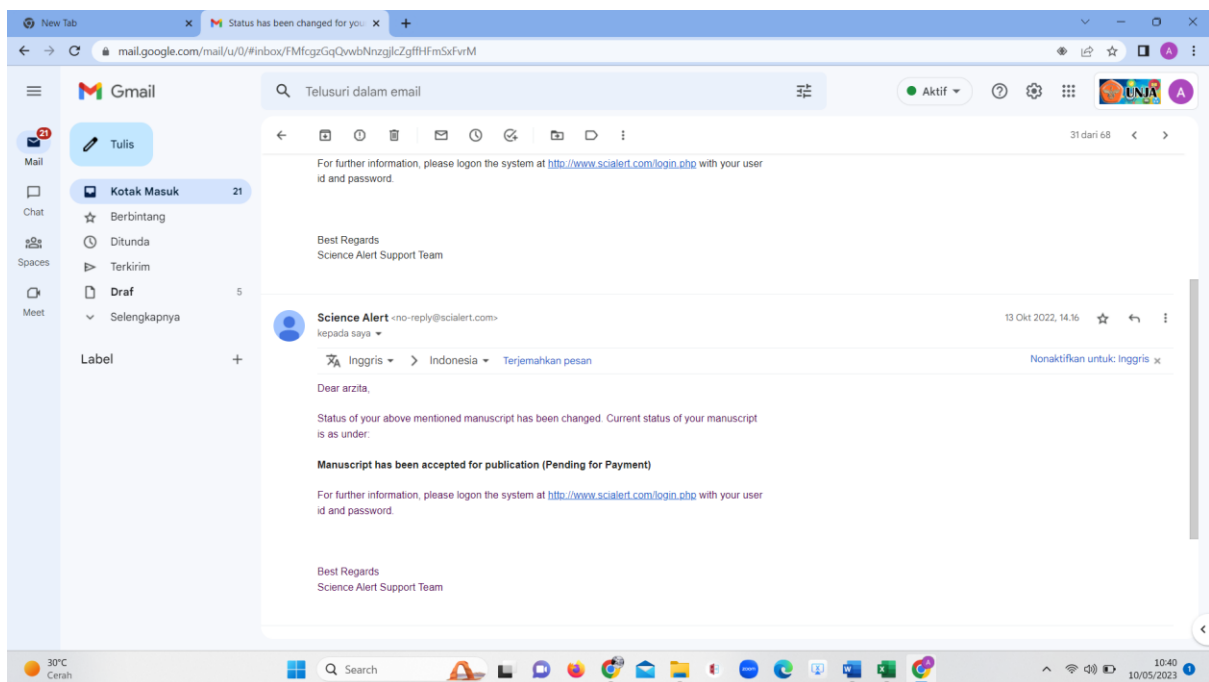
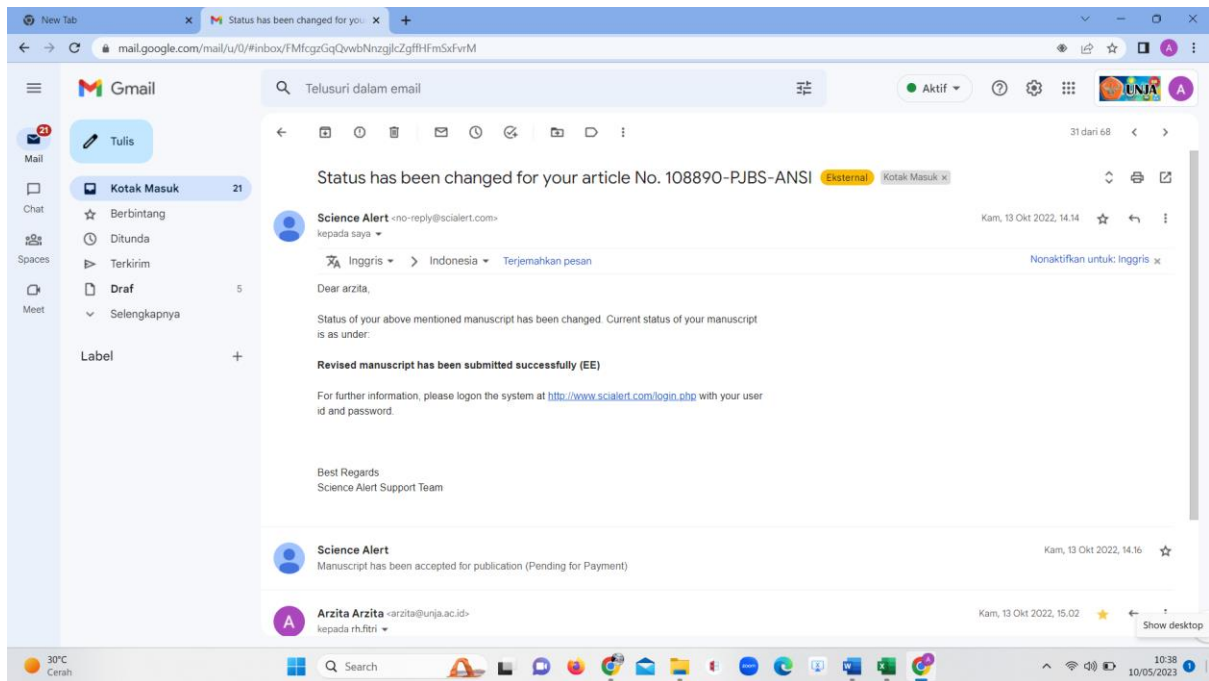










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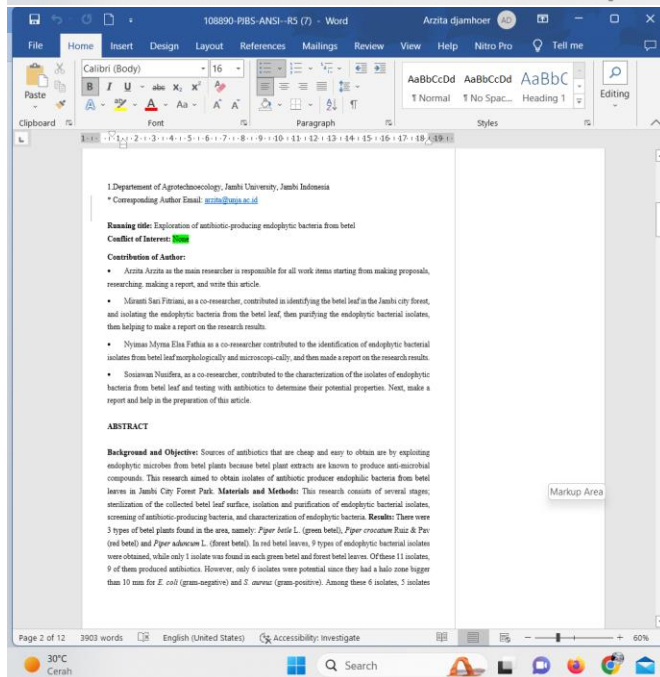
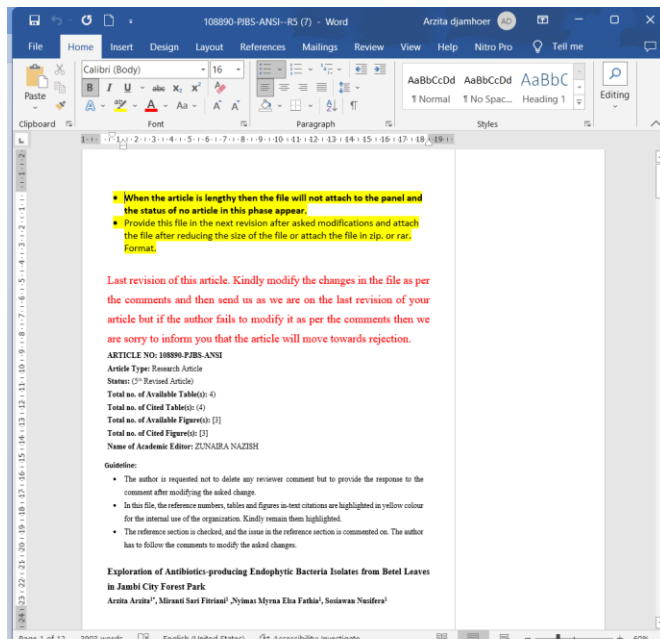
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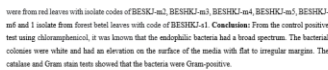
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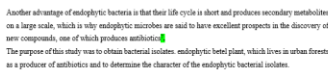




Keywords: Exploration, bacteria, endophytic, antibiotics, betel leaf, forest park, fermentation

INTRODUCTION

is a type of extracellular product, which is usually used as an alternative, as an antibiotic without any side effects. The leaf extract can produce anti-bacterial compounds, including carbonyl carboxylic and sulfide. Green benthic extracellular bacteria have antibacterial activity against fungi *Polyporus*.¹ The potential of endophytic bacteria in hotel food sources to produce new antibiotics is very good, so it is necessary to research the endophytic bacteria in hotel food sources. The endophytic bacteria in hotel food sources are the type of endophytic bacteria in hotel plants are highly dependent on the hotel food sources they grow in. So in this study, the location of the hotel food sampling was determined in the Jamnichi city forest park, known as the Muhammadiyah Jamnichi City Forest Park, in Kendal Village, Ampel District, North Jember City, Jember Regency, East Java Province, Indonesia. There are 200 species of plants, trees and shrubs, with the presence of several benthic plants, it is possible to find endophytic bacteria that are different from other endophytic bacteria, because climate, nutrition and the type of plant are different. The endophytic bacteria in hotel food sources are the type of endophytic bacteria that are resistant to antibiotics are quite abundant, this also fits the need for new and more effective antibiotics. The production of antibiotics can be done by chemical synthesis processes, from microbes and plants. The manufacture of antibiotics in Indonesia is highly dependent on imports of medicinal raw materials, now the import value is still high (90%). In effect to reduce imports, the government has been encouraging the manufacture of antibiotics in Indonesia. The manufacture of antibiotics, the production of which is through fermentation.² Research shows, Endophytic microbes that are a lot of attention because there are still many that have not been characterized, so their potential has not been maximized, especially in our country, Indonesia.³



MATERIALS AND METHODS

Time and Place

This research has been carried out at the UPT Basic and Central Laboratory, Andalas University, Padang, and the Faculty of Microbiology Laboratory Saintek, Jambi University, Mendalo Indah Village, Subdistrict Jambi Out of Town, Regency Muaro Jambi from June - November 2021.

Tools and Materials

Research using equipment; laminar flow, oven, incubator, autoclave, microscope, microwave, petri dish, test tube, Erlenmeyer, glass beaker, micropipette, glass, loop needle, knife, scalpel, pH meter, thermometer, micrometer, caliper, and analytical balance.

The materials used are: medium Nutrient Agar, Bacto Agar, Nutrient Broth, Tryptic Soy Broth, colouring agents, alcohol, Lugol's, distilled water, coffee paper, labels, permanent markers, and chemicals for characterization of endophytic bacteria, aluminium foil, and betel leaf.

Sampling

A sampling of leaves from the three types of betel plants studied: (green betel, red betel, and forest betel) was conducted by purposive random sampling method, samples were taken from the location Jambli City Forest Park area. Leaf samples were taken that looked healthy, not young leaves or old leaves, each leaf sample was taken with as many as ten leaves from each cluster.

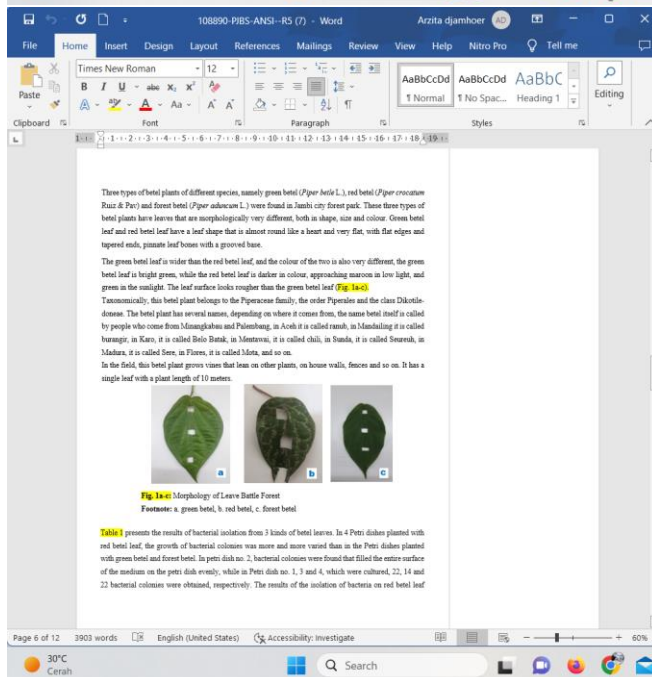
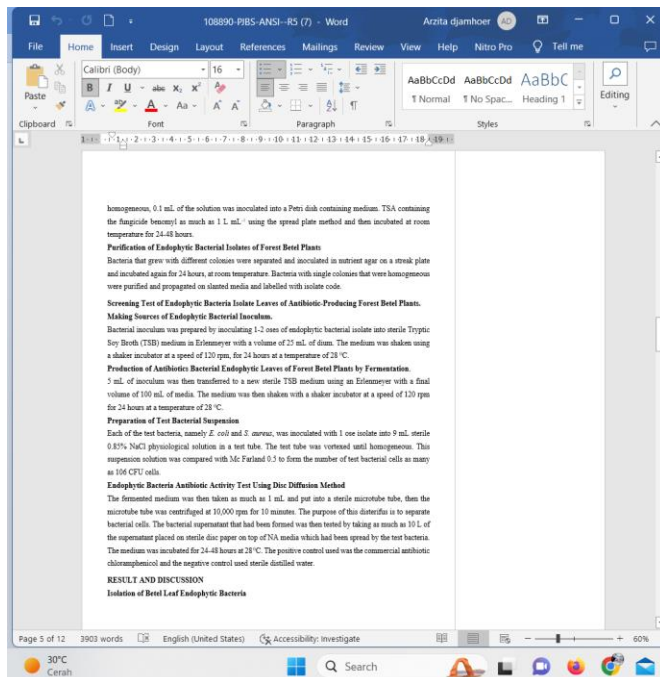
sample was taken with as many as ten leaves

Sterilization of Surface Forest Betel Leaf

The surface of the forest betel leaf that has been taken is then carried out by surface sterilization by immersing each forest betel leaf into sterile distilled water for 10 minutes, then transferred to the second solution, namely 70% ethanol solution for 10 minutes, then rinsed with sterile distilled water for 5 minutes. The surface of the sterilized betel leaf is then cut off the leaf part near the leaf base with a size of 1 x 1 cm using a sterile scalpel.

Isolation and Purification of Jambi Betel Leaf Endophytic Bacteria

For isolation and purification of endophytic bacteria, sterile plant segments were cut into pieces, ground until smooth, then put into a 0.85% physiological NaCl solution and homogenized, after being



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found 9 different types of bacteria, while on green betel leaf and forest betel, each of them found only 1 type of bacteria from 4 cultured Petri dishes.

Table 1. Isolation Results of Endophytic Bacteria from Forest Betel Leaf Plants

No	Betel type	Point	Number of colonies	Purified isolate
1.	Green betel (<i>Piper hirtell</i> L.)	I	4	
		II	4	
		III	5	
		IV	11	3
2.	Red betel (<i>Piper croceum</i> Rostk & Schmidt)	I	22	3
		II	Full spread	1
		III	14	2
		IV	22	3
3.	Forest betel (<i>Piper aduncum</i> L.)	I	6	
		II	Full	
		III	Full	
		IV	6	

The number of colonies that live from each type of betel leaf is different as shown in Table 1. A large number of bacterial colonies were obtained on a red betel leaf. The interaction of endophytic bacteria and plants is a form of symbiosis. The symbiosis between plants and endophytic bacteria is mutual, mutualism or commensalism. Mutualism symbiosis between endophytic bacteria and plants, in this case, endophytic bacteria get nutrients from plant metabolites and protect plants against the pathogen, while plants themselves obtain nutritional derivatives and active compounds needed for life.

The endophytic bacterial colonies isolated from these forest plants showed diversity, both in terms of colour, shape and growth speed. This was following which stated that endophytic bacteria in one host plant generally consist of several genera and species. The diversity of endophytic bacteria in a plant is also influenced by plant growth conditions, especially soil conditions. In some cases, plants with the same type or species have endophytic bacteria that were not always the same. In some plants, there are specific and distinctive endophytic bacteria that inhabit these plants.

Screening of Endophytic Bacteria Isolates in Forest Betel Leaf Produce Antibiotics

11 different endophytic bacterial isolates were then tested for screening as producers of antibiotics, were presented in Figure 2 and Table 2.

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Figure 2 shows isolates of endophytic bacteria from 3 kinds of betel leaf from Jambi city forest

Table 2. The results of the screening test for endophytic bacterial isolates from 3 types of betel leaf from Jambi City Forest Park

No	Sample Name	Origin of isolate	Isolate Code	Inhibition Zone d (mm)	d, average (mm)	Screening Isolate Bacteria	Test
1.	Green betel (<i>Piper hirtell</i> L.)	20/1	1	BB10087-1-1	0	0	No activity
		21/1	2	BB10087-1-2	0	0	No activity
		22/1	3	BB10087-1-3	11	10	No activity
		23/1	4	BB10087-1-4	20	20	No activity
2.	Red betel (<i>Piper croceum</i> Rostk & Schmidt)	20/2	5	BB10087-2-5	37	20/2	No activity
		21/2	6	BB10087-2-6	11/3	11/3	No activity
		22/2	7	BB10087-2-7	11	11	No activity
		23/2	8	BB10087-2-8	10/3	10/3	No activity
		24/2	9	BB10087-2-9	0	0	No activity
		25/2	10	BB10087-2-10	0	0	No activity
		26/2	11	BB10087-2-11	0	0	No activity
		27/2	12	BB10087-2-12	0	0	No activity
3.	Forest betel (<i>Piper aduncum</i> L.)	21	13	BB10087-3	14/20	14	No activity
4.	Positive Control (<i>Klebsiella pneumoniae</i>)			10	21/3	10	No activity
5.	Negative control (sterile water)			0	0	0	No activity

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Table 1 and Figure 2 above can be seen that 11 isolates had activity as antibiotics, which were able to kill *E. coli* and *S. aureus* bacteria. Of these 11 isolates, six potential isolates were obtained, namely the activity of the inhibition zone above 10 mm. Six of these isolates are indicated by the isolated code BESHRJ-m-2, BESHRJ-m-3, BESHRJ-m-4, BESHRJ-m-5, BESHRJ-m-6 dan BESHRJ-m-1. The six isolates of endophytic bacteria are categorized as strong inhibitors because they have inhibition above 10 mm (Fig. 3a). In Figure 3b, isolates of endophytic bacteria No. 6-X, forming a halo zone that is larger than the other isolates, Figure 3b-Y. The halo zone formed is larger than the halo zone formed by the antibiotic Chloramphenicol because of the positive control (Figure 3b-Z).

Figure 3a and 3b

Figure 3a shows the killing power of antibiotics and endophytic bacteria from 3 kinds of betel leaf from Jambi city. Figure 3b shows the killing power of antibiotics and endophytic bacteria from 3 kinds of betel leaf from Jambi city.

Figure 3a: Endophytic bacteria, 3b-Y, clear zone, 3b-Z, chloramphenicol positive control.

Figure 3b: Endophytic bacteria, 3b-Y, clear zone, 3b-Z, chloramphenicol positive control.

The formation of a clear area around the colony of endophytic bacterial isolates indicates the possibility of antimicrobial compounds capable of killing or at least inhibiting the growth of pathogenic bacteria. This was confirmed by Indrawati et al. and Sijar et al. who conducted a similar test with endophytic bacteria from

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Indonesian medicinal plants. Isolated endophytic bacteria showed inhibitory activity against pathogenic bacteria characterized by the formation of a clear area around the endophytic bacterial colonies. These six potential endophytic isolates were categorized as broad spectrum because they could kill Gram-negative (*E. coli*) and Gram-positive (*S. aureus*) bacteria. The presence of the silvery zone produced from endophytic bacteria was because endophytic bacteria have antimicrobial compounds. The advantage of using these endophytic bacteria have the same compounds as the original plant. Betel plants contain phenolic compounds and their derivatives which can inhibit the growth of *Proteus* bacteria. The antibacterial mechanism of phenol compounds in killing microorganisms is by denaturing bacterial cell proteins and the presence of these compounds so that endophytic bacteria produce an inhibitory zone against the test bacteria *E. coli* and *S. aureus*.

Macroscopic and Microscopic Characterization of Isolates of Potential Endophytic Bacteria in Antimicrobial Producing Forest Betel Leaf

Macroscopic and microscopic characterization of potential isolates is presented in Table 3 and Table 4.

Table 3: Macroscopic Characterization of Potential Endophytic Bacteria of L. new Betel Forest

No	Sample Code	Isolate Code	Colour	Shape	Margin	Elevation
1	Na SM-1-2	BESHRJ-m2	White	Circular	Undulate	Convex
2	Na SM-1-3	BESHRJ-m3	Cream	Bird	Beveled	Convex
3	Na SM-1-4	BESHRJ-m4	White	Irregular	Undulate	Convex
4	Na SM-1-5	BESHRJ-m5	Cream	Irregular	Undulate	Convex
5	Na SM-1-6	BESHRJ-m6	White	Irregular	Undulate	Convex
6	Na SR-2-1	BESHRJ-1	Cream	Irregular	Undulate	Flat

Table 4: Microscopic Characterization of Isolates of Endophytic Bacteria Producing Antimicrobial Potential From Forest Betel Leaf

No	Sample Code	Isolate Code	Gram	Cell Shape	Arrangement	Endospore
1	Na SM-1-2	BESHRJ-m2	Positive	Bacilli	Diplotherial	Have
2	Na SM-1-3	BESHRJ-m3	Positive	Bacilli	Diplotherial	Have
3	Na SM-1-4	BESHRJ-m4	Positive	Bacilli	Streptothelial	Have
4	Na SM-1-5	BESHRJ-m5	Positive	Bacilli	Streptothelial	Have
5	Na SM-1-6	BESHRJ-m6	Positive	Bacilli	Streptothelial	Have
6	Na SR-2-1	BESHRJ-1	Negative	Bacilli	Streptothelial	Have not

Of these six potential isolates, it was found that they were very diverse based on their morphological characteristics. Meanwhile, on microscopic observation, five isolates were Gram-positive, one was Gram-

